

## THE ANTHROPOGENIC IMPACT ON THE CHEMICAL AND MICROBIOLOGICAL PROFILE OF THE PASĂREA RIVER, ROMANIA

**PODOSU (VLAD) Aurelia, NEAGU Simona, LUCACI Anca Ioana, RUGINESCU Robert, COJOC Roxana, BĂTRÎNESCU-MOTEAU Costin, PURCĂREA Cristina, ENACHE Mădălin**

**Abstract.** This study reported the chemical and microbiological composition of a sector of Pasărea River, Romania, comprising the Dimieni Bridge (DB) and Tunari Dam (TD) sites, in relation with the chemical and microbiological pollution of this tributary of River Dâmbovița. The amount of sulfate-reducing bacteria ( $5.75 \times 10^7$  cells/mL at DB and  $5.5 \times 10^7$  at TD), nitrite and nitrate bacteria ( $3 \times 10^3$  cells/mL at DB and  $3 \times 10^3$  at TD), total coliforms ( $2.5 \times 10^3$  C.F.U./mL at DB and  $2.25 \times 10^3$  at TD) and faecal streptococci ( $5 \times 10^3$  C.F.U./mL at DB and  $1.4 \times 10^2$  at TD) revealed a relatively high cell density of microbial groups considered indicators of microbiological pollution. The analysis of the chemical composition highlighted the presence of chromium, cerium, aluminium and barium oxides in the 10 – 20 % range at DB in winter and in the 20 – 70% range at TD during spring confirming the chemical pollution of the analysed river sector. These contents were in accordance with values that could sustain the development of a polluting microbial community. Based on the corroborated data, the current survey proposed a set of criteria for establishing standards for the microbiological and chemical pollution of fresh water relative to drinking water values.

**Keywords:** Pasărea River, polluted water, microbial and chemical composition.

**Rezumat. Impactul antropogen asupra produselor chimice și microbiologice ale râului Pasărea, România.** Acest studiu a prezentat compoziția chimică și microbiologică a unui sector al râului Pasărea, România, cuprinzând siturile Podul Dimieni (DB) și Barajul Tunari (TD), în relație cu poluarea chimică și microbiologică a acestui afluent al râului Dâmbovița. Cuantificarea bacteriilor sulfato-reducătoare ( $5,75 \times 10^7$  celule/mL la DB și  $5,5 \times 10^7$  la TD), bacterii nitriți și nitrați ( $3 \times 10^3$  celule/mL la DB și  $3 \times 10^3$  la TD), coliforme totale ( $2,5 \times 10^3$  C.F.U./mL la DB și  $2,25 \times 10^3$  la TD) și streptococii fecali ( $5 \times 10^3$  C.F.U./mL la DB și  $1,4 \times 10^2$  la TD) au relevat o densitate celulară relativ mare a grupelor microbiene considerate indicatori ai poluării microbiologice. Analiza compoziției chimice a evidențiat prezența oxizilor de crom, ceriu, aluminiu și bariu în intervalul 10 – 20 % la DB iarna și în intervalul 20 – 70 % la TD în timpul primăverii confirmând poluarea chimică a sectorului fluvial analizat. Aceste conținuturi erau în concordanță cu valori care ar putea susține dezvoltarea unei comunități microbiene poluante. Pe baza datelor coroborate, studiul actual a propus un set de criterii pentru stabilirea standardelor de poluare microbiologică și chimică a apei dulci în raport cu valorile apei potabile.

**Cuvinte cheie:** Râul Pasărea, apă poluată, compoziție microbială și chimică.

### INTRODUCTION

The biological activity of any type of ecosystem would not be possible without the existence of the microbial component. This establishes the balance between organic and inorganic substances, respectively between living and non-living matter. Flowing waters represent a particular environment from a microbiological point of view because their microbial composition undergoes fundamental changes from the springs to the confluence of the river with other flowing waters, or with the river-sea system (McCABE, 2011; KRAUSE et al., 2022). This composition is influenced by the hydrographic basin, by the input of the tributaries, the physical-chemical and climatic conditions, but mostly by the anthropogenic impact. Human communities have developed in the proximity of water reserves (flowing waters, lakes, seas, oceans) with a significant impact on them (BURIAN et al., 2000). Residue discharges resulting from industrial, agricultural, recreational or other human activities produce significant changes in the physical-chemical and microbiological composition of water (SINTON et al., 1993; HOLCOMB & STEWART, 2020). Water plays an essential role for all life forms on Earth, its quality having a major impact on all human communities. Throughout history, human communities have developed in the vicinity of rivers using water resources for various activities, which has led to an increase in anthropogenic impact on water quality (MILLER et al., 2017). This composition in turn influences the community of microorganisms in the anthropized aquatic ecosystem with direct effects on the cyclical rhythm of the biogeochemical cycles (ABBOTT et al., 2018).

In general, there is no system based on which water could be considered polluted or unpolluted, the classification being based on certain human perceptions. For example, water is considered sweet, brackish or salty according to taste, a threshold of 3g/L NaCl being arbitrarily chosen to separate a sweet water from a salty one (ENACHE et al., 2017). There are normative systems that classify water into several quality classes, a classification that allows the use of water in certain types of activities. Arbitrarily, in this study it was considered that the investigated running water can be considered polluted or unpolluted depending on the physical, chemical and microbiological parameters established for the quality of drinking water provided for the socio-economic systems along the investigated area.

The microbiological component is essential for the functionality of an ecosystem (aquatic, terrestrial) in terms of flow of matter, energy, information and biogeochemical cycles (AZAM et al., 1983). Changes in the physical and chemical composition of the ecosystem are reflected in its microbiological activity. Water reserves are of fundamental importance for daily human activities, so that knowledge of their biological and chemical structure is of particular importance. In the present study, a sector of the Pasărea River was investigated, located in the vicinity of some fundamental economic units for the normal development of daily activities, but also of a strongly developed human

community with a significant anthropogenic impact on this sector of the river. The Pasărea River (Fig. 1) is part of the Argeş River hydrographic basin, being a typical lowland river with its sources in the Otopeni forest at an altitude of 91 meters, crossing the city of Bucharest in the northeast. This river merges with the Dâmbovița river (in the town of Fundeni), at an altitude of 40 meters, after a course of 48 km, with a meandering course with a sinuosity coefficient of 1.5 and after collecting the waters of the Sindrilița tributary (11 km long) with which it drains an area of 254 km<sup>2</sup>. The meandering course makes it difficult for water to drain and favors the appearance and development of swamp vegetation. Water runoff is defined by average annual flows of less than 1m<sup>3</sup>/s, the speed of the water current being low as a result of the low slopes. In this regard, the river bed was modified, according to the sector investigated in this study, with 24 ponds used for fish farming and irrigation, including Tunari, with an area of 21 ha (COCOȘ, 2006).

The study also aims to establish some general criteria by which a sector of a flowing water can be considered polluted or not, considering the biological capacity of a river to recover qualitatively as a result of the biological activity closely correlated with the physical and chemical composition.

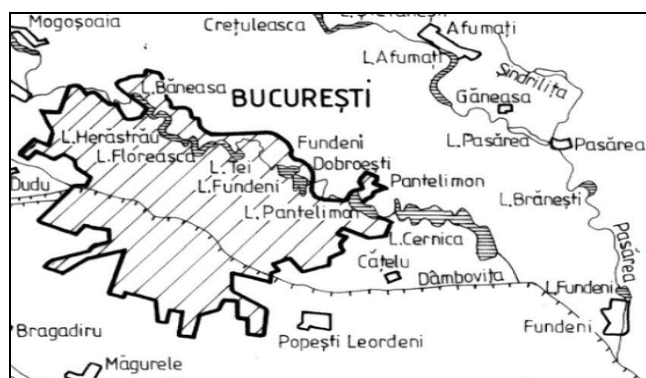


Figure 1. The location of the Pasărea River in the hydrographic basin of Bucharest area (COCOȘ, 2006).

## MATERIALS AND METHODS

*Water samples* were collected seasonally during summer, autumn and winter of 2020, and spring of 2021 from two locations identified as the Dimieni Bridge (DB) and the Tunari Dam (TD).

A series of *physical and chemical parameters* of the river water was measured *on site* counting pH, salinity, total dissolved solids (ppm), conductivity 1 (μS/cm), conductivity 2 (μS/cm<sup>2</sup>), salinity (PSU), total dissolved oxygen (mg O<sub>2</sub>/L), turbidity (NTU), water temperature (°C).

*The chemical composition* of water samples was determined by X-ray fluorescence spectrometry, using WD-XRF (Rigaku, Japan) equipment (NEAGU et al., 2021).

*Microbiological analyses* were performed from the samples and decimal dilutions for the main groups of microorganisms involved in the carbon, nitrogen, sulphur circuits as well as the groups of microorganisms indicating pollution following culture media and growth conditions as described in a previously papers (\*\*\*. APHA 1998; LUCACI et al., 2019; NEAGU et al., 2021). The composition of culture media was used for different groups of microorganisms corresponding for ammonifying, denitrifying, coliforms, heterotrophic, streptococcus and sulphate reducing bacteria (\*\*\*. APHA 1998; BERNASCONI et al., 2003; JANELIDZE et al., 2011).

*Pollution criteria* for flowing water were proposed relative to standards for drinking water (\*\*\*. CD - 98/83/EC; \*\*\*. STAS 3001-91) and previous data (MESSLEY & KINGSBURY, 1973; SINTON et al., 1998; RAJI et al., 2015; SELVAM et al., 2015; WANG et al., 2016; HOLCOMB & STEWART, 2020; NEAGU et al., 2021; PODOSU et al., 2021a, b; BĂTRÎNESCU-MOTEAU et al., 2022; RUGINESCU et al., 2022).

## RESULTS AND DISCUSSIONS

*The physical and chemical parameters of Pasărea River water samples.* The areas selected for the study, being controlled by the Pasărea river dam system, varied in terms of physical and chemical parameters.

This control system is reflected in significant values for turbidity, conductivity and total dissolved solids (DB – Fig. 2 and TD – Fig. 3). Indirectly, the rest of the parameters investigated on site were influenced by this system, as a positively correlated factor with turbidity and dissolved oxygen, for example. Regarding the spatial and temporal characteristics of the parameters, factors that varied significantly between ecosystems and factors that varied seasonally were highlighted. Similar behaviors were also reported in some studies for Georgian Black Sea coastal ecosystem (JANELIDZE et al., 2011).

*Water temperature* varied between 4 °C for the Dimieni Bridge and 7 °C for the Tunari Dam in winter and 29 °C in summer for both sites (Figs 2; 3), indicating small variations between the two locations. These moderate temperatures in the range of the tolerance limits for metabolic processes (LAZĂR et al., 2017) could sustain the development of microbial communities during all seasons.

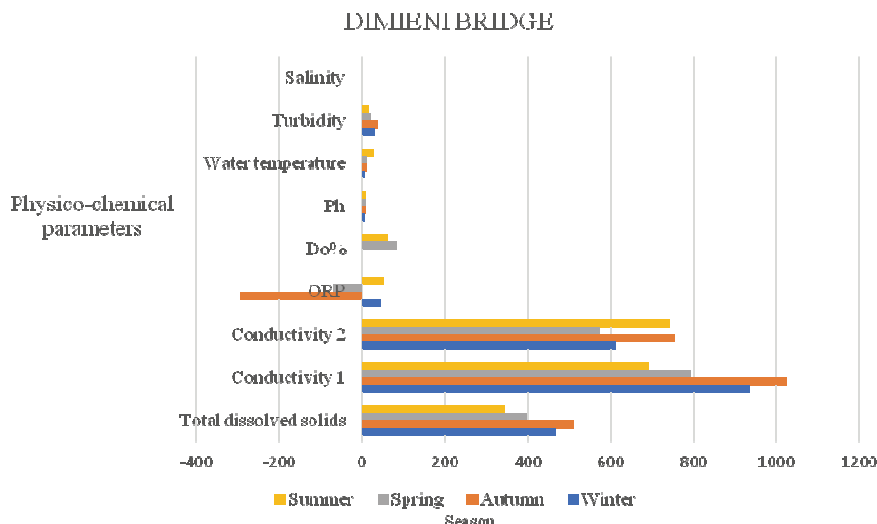


Figure 2. Parameters for the Dimieni Bridge: Total dissolved solids (ppm); Conductivity 1 ( $\mu\text{S}/\text{cm}$ ), Conductivity 2( $\mu\text{S}/\text{cm}^2$ ); Salinity (PSU); Total dissolved oxygen ( $\text{mg O}_2/\text{L}$ ); Turbidity (NTU); Water temperature ( $^\circ\text{C}$ ).

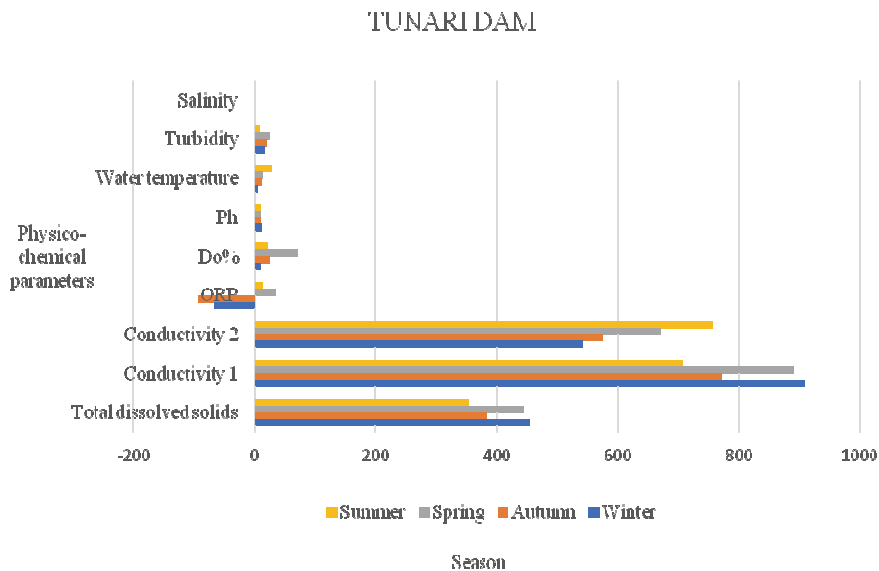


Figure 3. Parameters for the Tunari Dam.

The pH values of the water samples from the Dimieni Bridge site (Fig. 2) ranged between 6.82 in the winter season and 8.44 in the summer season, while for the Tunari Dam, the recorded values were higher, in a slightly alkaline range, varying between 8.27 during summer season and 9.55 in winter (Fig. 3). These relatively limited variations around a neutral pH during winter times have minor bactericidal effect by protein denaturation of hydrolysis, while the more alkaline values (8.44 and 9.55, respectively) could be responsible for modelling de composition of the river microbiome (LAZĂR et al., 2016).

*Oxid Reducing Potential (ORP).* The ORP measuring the electrical charge of the water revealed negative values for all analysed samples (Figs. 2 and 3), indicating an antioxidant potential that contributes to the neutralization of water acidity. As a parameter correlated with pH values, negative ORP values corresponded to higher pH values observed during autumn in both locations. Thus, the ORP value for the Dimieni Bridge (-293 mV) was in accordance with the higher pH value (8.27) at 11 °C (Fig. 2), while for the Tunari Dam, the negative ORP (-92 mV) correlated with a higher pH value (9,16). Moreover, during winter season, the negative ORP (-66.7 mV) value is supported by the higher pH value (9,55) (Fig. 3).

*Dissolved oxygen (O<sub>2</sub>%).* The oxygen saturation (DO%) measured in the two locations of the Pasărea River was highly different, varying also with the season. At the Tunari Dam, the DO content ranged from 9.2% in winter to 70.6% in spring (Fig. 3), while at the Dimieni Bridge the DO was 0 during both winter and autumn seasons, and varied between 62.2-83.5 in summer and spring (Fig. 2). The measured oxygen content of the river water in the mass of water was in accordance with supporting the development of aerobic microorganisms. Moreover, these recorded DO values

corroborated with those of the corresponding oxid reducing potential ones (Figs. 2 and 3) could explain the absence of microorganisms involved in the nitrogen biogeochemical cycle, considering diminishes nitrate processing in the anthropized water flow (AZIZIAN et al., 2017).

*The microbial content of the Pasărea River.* During the four seasons, the number of microbial colony-forming units/ml obtained from water samples collected from the two locations of the Pasărea River showed a variation with both the type of microorganisms and the season (Figs. 4 and 5). In the case of the Dimieni Bridge, these values ranged from  $3.22 \times 10^5$  (summer) to  $4.5 \times 10^4$  (spring) (Fig. 4). For the sample, from the Tunari Dam, considering figure 5, the highest number of colony-forming units/ml was highlighted in the sample taken in the winter season,  $1.7 \times 10^5$  and recorded a decrease in the spring season, respectively  $1.4 \times 10^2$  CFU/ml. In temperate zones, the development of heterotrophic bacteria is dependent on the seasonal temperature variations, while low temperatures inhibit microbial growth (POMEROY & WIEBE, 2001). In this context, the data obtained for the Dimieni Bridge water samples suggest the constant presence of the food source in all four seasons. For the Tunari Dam, the highest microbial content ( $1.7 \times 10^5$  UFC/ml) was recorded during winter season, in accordance with the high TDS value (454 ppm) indicating an uptake of organic matter of exogenous origin.

The presence of total coliforms, faecal coliforms and faecal streptococci is frequently associated with the faecal-household contamination of ecosystems (KOLAREVIC et al., 2011). In this study, the presence of total coliform bacteria was confirmed during the four seasons for the water sample from the Dimieni Bridge, the highest number of CFU/ml ( $6.2 \times 10^3$ ) being determined for samples collected in autumn (2020). The number of UFC/ml was constant during the following winter ( $1.85 \times 10^3$  UFC/ml) and spring ( $1.8 \times 10^3$  UFC/ml).

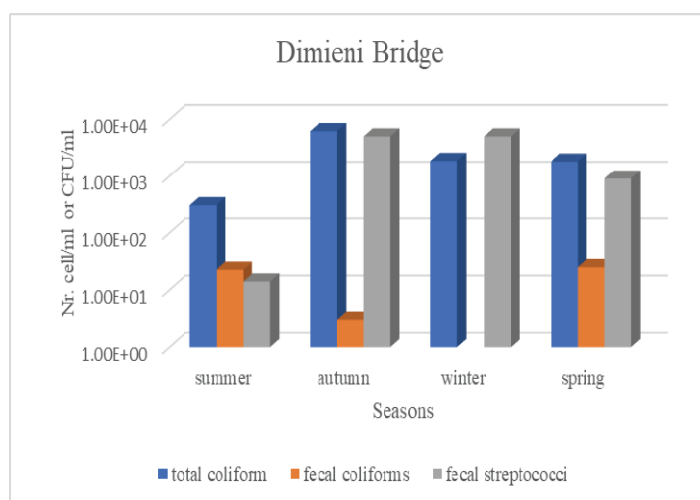


Figure 4. Microbial content of the Pasărea River during various seasons. The cell density (CFU/ml) of total coliforms, fecal coliforms and fecal streptococci was determined as indicated in Methods for water samples collected from the Dimieni Bridge and the Tunari Dam.

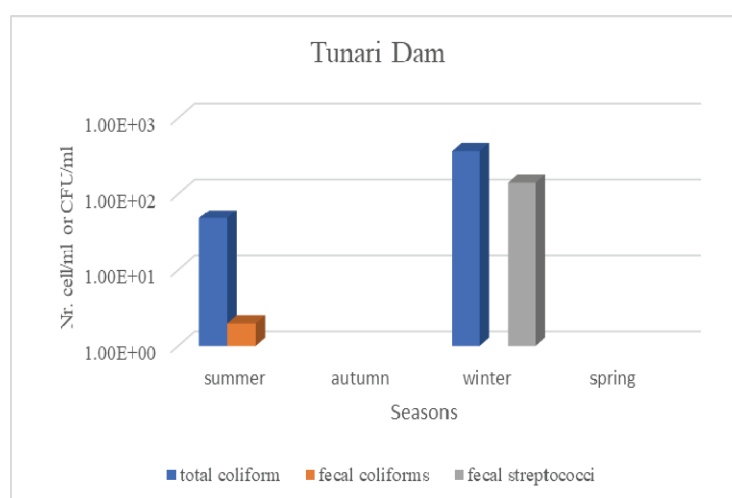


Figure 5. Groups of microorganisms, pollution indicator for the Tunari Dam.

A slightly reduced content of total coliforms, ( $3.03 \times 10^2$  CFU/ml), was determined in the water samples collected during the warm season (summer 2020). For the Tunari Dam samples, the specific faecal coliforms and streptococcal colonies were present only in the summer season (2020) with a recorded cell density of  $4.8 \times 10$  UFC/ml that increased 10-fold ( $3.65 \times 10^2$  UFC/ml) during winter (2021). For the samples collected in autumn (2020) and spring (2021), no total coliform bacteria were identified (Figs. 4 and 5)

Faecal coliform bacteria were scarcely present during the seasonal sampling, for the water sample from the Dimieni Bridge in winter ( $2.5 \times 10$  UFC/ml), summer ( $2.25 \times 10$  UFC/ml), and autumn (3 UFC/ml) seasons, while absent during winter. The Tunari Dam water samples revealed the presence of only 2 UFC/ml faecal coliform bacteria during summer season, and absence of these pathogens during the other seasons. The faecal streptococci content at the Dimieni Bridge varied depending on the season, ranging from  $5 \times 10^3$  UFC/ml during autumn and winter to  $1.4 \times 10$  UFC/ml in summer, with an intermediate value ( $9 \times 10^2$  UFC/ml) during spring (Fig. 4). Meanwhile, for the Tunari Dam location, faecal streptococci were recorded only during the winter season ( $1.4 \times 10^2$  UFC/ml) (Fig. 5).

Sulphate-reducing bacteria are an indicator of ecosystem pollution with organic substances, being able to reduce the microbial biomass due to the accumulation of  $H_2S$  toxic for aquatic organisms (LAZĂR et al., 2017). The high content of sulphate-reducing bacteria recorded in water samples from the Dimieni Bridge (Fig. 6) suggested a high anaerobic activity for organic matter degradation, with maximum ( $5.75 \times 10^7$  cells/ml) reached during the spring period. For the Tunari Dam (Fig. 7), the cell density of aerobic heterotrophs varied between  $7.5 \times 10^5$  cells/ml in winter and  $2.25 \times 10^5$  cells/ml during summer season, with  $10^2$ - $10^3$ -fold lower values during autumn and spring, respectively. At this site, the number of sulphate-reducing bacteria registered the highest content during the winter season ( $5.5 \times 10^7$  cells/ml), with a large drop during summer ( $5.5 \times 10^4$  cells/ml) and a lower presence in spring and autumn ( $4.75 \times 10^2$  cells/ml) (Fig. 7).

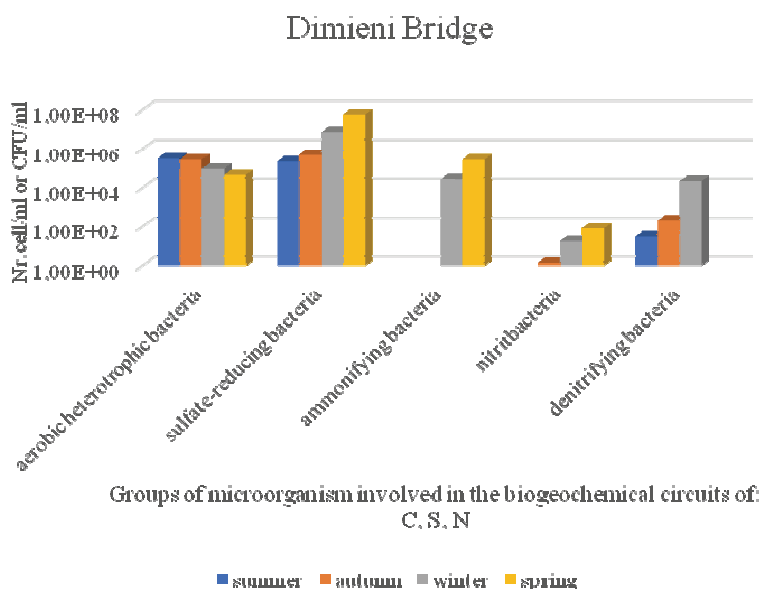


Figure 6. Groups of microorganisms involved in the biogeochemical circuits of carbon (C), nitrogen (N), sulphur (S) identified in the Dimieni Bridge site.

Bacteria involved in ammonification and nitrification (nitrite bacteria and nitrate bacteria) processes were present at the Dimieni Bridge site only in summer at a calculated content of  $3 \times 10$  cells/ml (Fig. 7), possibly related with the lowest content of total dissolved solids (345 ppm) recorded during this season (Fig. 2). During autumn, nitrite and denitrifying bacteria were identified at this River site at a 1.4 cells/ml and  $2.2 \times 10^2$  cells/ml content, respectively. The increased number of denitrifying bacteria could be justified by the highest content of total dissolved solids (510 ppm) recorded during autumn season (Fig. 2). Moreover, the absence of ammonifying bacteria and nitrate bacteria was also noted during this period of time (Fig. 7). For the same sampling point, an increased number of bacteria involved in ammonification ( $2.8 \times 10^4$  cells/ml), of nitrifying bacteria ( $1.8 \times 10$  cells/ml) and denitrifying bacteria ( $2.2 \times 10^4$  cells/ml) was identified during the winter season, while nitrate bacteria were absent. In spring, only ammonifying bacteria ( $2.8 \times 10^5$  cells/ml) and nitrite bacteria at low density ( $8 \times 10$  cells/ml) were identified among the nitrogen metabolizing microorganisms (Fig. 7).

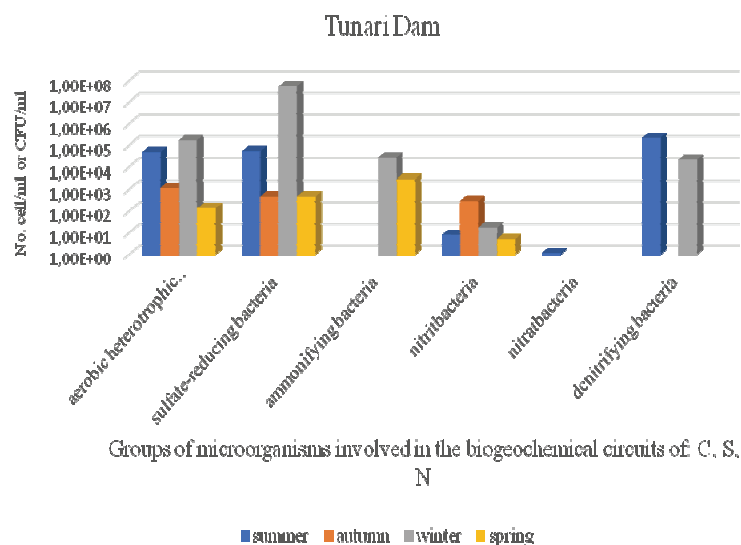


Figure 7. Groups of microorganisms involved in the biogeochemical circuits of carbon (C), nitrogen (N), sulphur (S) identified in the Tunari Dam site.

At the Tunari Dam, ammonifying bacteria were absent during the summer season, and only a low number of nitrite bacteria (8 cells/ml) and nitrate bacteria (1.18 cells/ml) were identified, while the denitrification process was highly present ( $2.2 \times 10^5$  cells/ml). A comparison of the seasonal microbial content involved in the nitrogen cycle indicated the dominance of denitrifying bacteria during summer ( $8 \times 10^4$ ), the sole presence of nitrite bacteria ( $3 \times 10^2$  cells/ml) during autumn, an intensification of the processes underlying the nitrogen biogeochemical circuit during winter represented by ammonifying ( $3 \times 10^3$  cells/ml), nitrite ( $1.8 \times 10^3$  cells/ml), and denitrifying ( $2.2 \times 10^4$  cells/ml) bacteria, and a conserved ammonifying bacteria content while reduced presence (5 cells/ml) of bacteria involved in the nitrification process during spring season. The increased number of bacteria identified in the three stages of the cold season could be due to the highest values of the TDS content that was recorded at this location during this year interval. The reported data suggests an incomplete biogeochemical nitrogen cycle occurring at the two locations of the Pasărea River (ZARNEA, 1994; ARDELEAN, 2012; LAZĂR et al., 2016). Moreover, the determined physical and chemical parameters of the water samples from these investigated sites suggested the presence of polluted factors with an inhibitory effect on specific enzymatic activities involved in nitrogen biogeochemical transformations.

*The chemical composition of the water in the Pasărea River.* The chemical composition of the investigated Dimieni Bridge and Tunari Dam zones of the Pasărea River (Fig. 8) showed a large variation based on the sampling season. During summer, both sites contained SiO<sub>2</sub> (15-18 % mass), and lower content of K<sub>2</sub>O, CaO, Fe<sub>2</sub>O<sub>3</sub>, As<sub>2</sub>O<sub>3</sub>, Cl, Al<sub>2</sub>O<sub>3</sub>, while >35% mass TiO<sub>2</sub> was present at the Tunari Dam, and 14% mass of Tb<sub>4</sub>O<sub>7</sub> and Lu<sub>2</sub>O<sub>3</sub> at the Dimieni Bridge. Water samples collected during autumn (November 2020) showed a low content of SiO<sub>2</sub>, K<sub>2</sub>O, CaO, As<sub>2</sub>O<sub>3</sub> and Cl as compared to the summer season, with <10% in the water collected from the Tunari Dam, and < 5% at the Dimieni Bridge. In addition, 67% BaO and 3-6% of HO<sub>2</sub>O<sub>3</sub>, Gd<sub>2</sub>O<sub>3</sub> and Cr<sub>2</sub>O<sub>3</sub> were detected at the Dimieni Bridge, and 30% Nd<sub>2</sub>O<sub>3</sub>, 11% ReO<sub>2</sub>, 9.7% Fe<sub>2</sub>O<sub>3</sub>, 8.1% Tb<sub>4</sub>O<sub>7</sub> and 4.7% SrO at the Tunari Dam (Fig. 8).

During the winter season (February 2021), high concentrations of BaO (80% by mass) were identified at the Tunari Dam and Dimieni Bridge (35%), respectively, followed by lower mass percentages (<6) of Fe<sub>2</sub>O<sub>3</sub>, SiO<sub>2</sub>, CaO, K<sub>2</sub>O and Cl. Sm<sub>2</sub>O<sub>3</sub> (16% mass), Tb<sub>4</sub>O<sub>7</sub> (10% mass), Gd<sub>2</sub>O<sub>3</sub> (8% mass) determined at the Dimieni Bridge (Fig. 8).

The chemical composition during spring (April 2021) revealed an increase of the BaO mass percentage (73%) at the Dimieni Bridge, while only 20% at the Tunari Dam, as in the other seasons, and lower percentages (<5%) of SiO<sub>2</sub>, K<sub>2</sub>O and CaO. Water samples from the later location also showed a high content of CeO<sub>2</sub> (30%), followed by Fe<sub>2</sub>O<sub>3</sub> (8.6%), As<sub>2</sub>O<sub>3</sub> (6.12%), GeO<sub>2</sub> (4.85%) and SeO<sub>2</sub> (3.4%) (Fig. 8).

*Microbial and chemical criteria for the polluted water of the Pasărea River.* Based on current standard for drinking water quality (\*\*\*. Law 458/2002), and previous studies of water pollution (SELVAM et al., 2015; NEAGU et al., 2021), the current data were interpreted in the frame of criteria for polluted flowing water considering the presence of (i) sulphate reducing bacteria, nitrite and nitrate bacteria independent of their content, and (ii) Cr, Ce, Al and Ba oxides or salts at various concentration, including traces. Meanwhile, the presence of total coliforms and fecal streptococci confirmed that the analysed water sample was microbiologically polluted. Regarding the pollution origin, a ratio >1 between fecal coliform bacteria and fecal streptococci indicated anthropogenic pollution, while values <1 corresponded to animal pollution (MESSLEY & KINGSBURY, 1973; SINTON et al., 1998; RAJI et al., 2015; MILLER & HUTCHINS, 2017). For the Pasărea River, the corroboration of the presence of these factors indicated a highly polluted water sample, while the partial occurrence of these factors corresponded to a slow or moderate pollution of the flowing water.



Figure 8. Season variation of the chemical composition of the water in the Pasarea River at DB and TD sites.

## CONCLUSIONS

The physical and chemical parameters of the Pasărea River water indicated a variable pollution degree in two analysed sectors depending on the season. Suspended materials present at both sampling sites during the four seasons showed an increase in the electrical conductivity, with the highest values recorded at the Dimieni Bridge during autumn. The pH values ranged between 6.82 (winter - Dimieni Bridge) and 9.55 (winter - Tunari Dam), slightly exceeding the limits established in Order 161/16.02.2006, and suggesting an anthropogenic impact in the investigated areas.

The chemical composition of water highlighted the presence of oxides of rare metals such as  $\text{Sm}_2\text{O}_3$ ,  $\text{Gd}_2\text{O}_3$ ,  $\text{Tb}_4\text{O}_7$ ,  $\text{CeO}_2$ ,  $\text{Nd}_2\text{O}_3$ ,  $\text{Lu}_2\text{O}_3$ ,  $\text{TiO}_2$ ,  $\text{Cr}_2\text{O}_3$ . Microbiological analyses revealed the presence of faecal streptococci and total coliforms during the four seasons in the sample of the Dimieni Bridge, and faecal coliforms during summer, autumn and spring. For the Tunari Dam, the microbial pollution was due to microbial groups comprised total coliforms and faecal coliforms during summer, and total coliforms and faecal streptococci in winter. Barium-based compounds were present at high concentrations, in support of chemical and biological pollution of the investigated areas.

## ACKNOWLEDGEMENTS

We thank the Romanian National Water Administration for granting sampling permission and ensuring access at sampling sites. This work was supported by the RO1567-IBB05/2020 and RO1567-IBB05/2021 projects (Romanian Academy).

## REFERENCES

- ABBOTT B. W., MOATAR F., GAUTHIER O., FOVET O., ANTOINE V., RAGUENEAU O. 2018. Trends and seasonality of river nutrients in agricultural catchments: 18 years of weekly citizen science in France. *The Science of the Total Environment*. Elsevier. Paris. **624**: 845-858.
- ARDELEAN I. I. 2012. *Microbiologie generală*. Edit. Ars Docendi. București. **2**. 270 pp.
- AZAM F., FENCHEL T., FIELD J. G., GRAY J. S., MEYER-REIL L. A., THINGSTAD F. 1983. The ecological role of water - column microbes in the sea. *Marine ecology progress series*. MEPS Publisher. London: 257-263.
- AZIZIAN M., BOANO F., COOK P. L. M., DETWILER R. L., RIPPY M. A., GRANT S. B. 2017. Ambient groundwater flow diminishes nitrate processing in the hyporheic zone of streams: Ambient groundwater and stream N-cycling. *Water Resources Research*. Wiley Press. London. **53**: 3941-3967.
- BĂTRÎNESCU-MOTEAU C., NEAGU SIMONA, LUCACI A. I., RUGINESCU R., MARIA G., COJOC ROXANA, PURCĂREA CRISTINA, PODOSU AURELIA, ENACHE M. 2022. Preliminary data concerning communities of microorganisms in a volcanic tuff endolytic habitat. *Oltenia. Studii și Comunicări. Științele Naturii*. Muzeul Olteniei Craiova. **38**(1): 168-173.
- BERNASCONI C., DAVERIO E., GHIANI M. 2003. Microbiology Dimension in EU Water Directives, EUROPEAN COMMISSION JOINT RESEARCH CENTRE. *Institute for Environment and Sustainability Inland and Marine Waters Unit*. Ispra Press. London: 1-54.
- BURIAN S. J., NIX S. J., PITT R. E., ROCKY D. S. 2000. Urban wastewater management in the United States: past, present, and future. *Journal Urban Technology*. Taylor & Francis Press. **7**: 33-62.
- COCOȘ O. 2006. Managementul apei în municipiul București, *Sistemele hidrografice București*. Edit. Ars Docendi. București: 68-103.
- ENACHE M., TEODOSIU GABRIELA, ITOH T., KAMEKURA M., STAN-LOTTER H. 2017. Halophilic microorganisms from man-made and natural hypersaline environments: physiology, ecology and biotechnological potential, In: Helga Stan-Lotter & Sergiu Fendrihan (Eds.). *Adaptation of Microbial Life to Environmental Extremes*. Springer. Wien New York. **2**: 201-226.
- HOLCOMB D. A. & STEWART J. R. 2020. Microbial indicators of fecal pollution: recent progress and challenges in assessing water quality. *Curr. Environ. Health Reports*. Wiley Press. London. **7**: 311-324.
- JANELIDZE N., JAANI E., LASKHI N., TSKHVEDIANI A., KOKASHVILI T., GVARISHVILI T., JGENTI D., MIKASHVIDZE E., DIASAMIDZE R., NARODNY S., OBISO R., WHITEHOUSE C.A., HUQ A., TEDIASHVILI G. 2011. Microbial water quality of the Georgian coastal zone of Black Sea. *Marine Pollution Bulletin*. Elsevier. Paris. **62**: 573-580.
- KOLAREVIĆ S., KNEZEVIC-VUKCEVIĆ J., PAUNOVIĆ M., TOMOVIĆ J. M., GAČIĆ Z., VUKOVIC-GACIC B. 2011. The anthropogenic impact on water quality of the river Danube in Serbia: microbiological analysis and genotoxicity monitoring. *Archives of Biological Sciences*. Publisher Inst. Bioloska Istrazivanja Sinisa Stankovic. Beograd. **63**(4): 1209-1217.
- KRAUSE S., ABBOTT B. W., BARANOV V., BERNAL S., BLAEN P., DATRY T., DRUMMOND J., FLECKENSTEIN J. H., VELEZ J. G., HANNAH D. M., KNAPP J. L. A., KURZ M., LEWANDOWSKI J., MARTI E., MENDOZALERA C., MILNER A., PACKMAN A., PINAY G., WARD A. S., ZARNETZKE J. P. 2022. Organizational Principles of Hyporheic Exchange Flow and Biogeochemical Cycling in River Networks Across Scales. *Water Resources Research*. Wiley Press. London. **28**(3): 1-25.
- LAZĂR V., MĂRUȚESCU L. G., CHIFIRIUC M. C. 2016. *Microbiologie generală și aplicată*. Edit. Universității din București. 408 pp.
- LAZĂR V., CURUTIU C., DITU L. M., HOLBAN A., GHEORGHE I., MARINESCU F., ILIE M., MARCU E., IVANOV A., DOBRE D., CHIFIRIUC M. C. 2017. Physico-Chemical and Microbiological Assessment of



- Organic Pollution in Plain Salty Lakes from Protected Regions. *Journal of Environmental Protection*. Scimago Press. London. **8**: 1474-1489.
- LUCACI A. I., MOLDOVEANU MIRELA, FLORESCU LARISA, COJOC ROXANA, NEAGU SIMONA, RUGINESCU R., ENACHE M. 2019. The seasonal dynamics of the cultivable microbial communities in Letea saline lake. *AgroliLife Scientific Journal*. Publications of the University of Agronomic Sciences and Veterinary Medicine of Bucharest. **8**(1): 160-166.
- MCCABE D. J. 2011. Rivers and Streams: Life in Flowing Water. *Nature Education Knowledge*. MDPI Press. London. **3**(10): 19.
- MILLER J. D. & HUTCHINS M. 2017. The impacts of urbanisation and climate change on urban flooding and urban water quality: a review of the evidence concerning the United Kingdom. *Journal of Hydrology: Regional Studies*. Cambridge University Publishing. London. **12**: 345-362.
- MESSLEY K. E. & KINGSBURY P. J. 1973. The Fecal Coliform/Fecal Streptococcus Ratio as a Measure of Bacterial Contamination and Indicator of Its Source in the Des Moines River. *Proceedings of the Iowa Academy of Science*. The Iowa Academy of Science Publishing. **80**(2): 74-77.
- NEAGU SIMONA, ENACHE M., COJOC ROXANA, RUGINESCU R., MOLDOVEANU MIRELA, FLORESCU LARISA, LUCACI I. 2021. Seasonal variation of the water color from the IOR lake – Bucharest. *Oltenia. Studii și comunicări. Științele Naturii*. Muzeul Olteniei Craiova. **37**(1): 205-210.
- PODOSU AURELIA, BĂTRÎNESCU-MOTEAU C., LUCACI I., RUGINESCU R., COJOC ROXANA, NEAGU SIMONA, PURCĂREA CRISTINA, FLORESCU LARISA, MOLDOVEANU MIRELA, ENACHE M. 2021a. The variation of physico-chemical parameters in different seasons in high polluted sectors of the Pasărea River. The Scientific International Conference “The Museum and Scientific Research” *Biodivest 2021*, Book of abstract. Muzeul Olteniei Craiova: 88.
- PODOSU AURELIA, NEAGU SIMONA, COJOC ROXANA, RUGINESCU R., LUCACI I., BĂTRÎNESCU-MOTEAU C., FLORESCU LARISA, MOLDOVEANU MIRELA, PURCĂREA CRISTINA, ENACHE M. 2021b. The seasonal variation of physico-chemical parameters in polluted rivers following anthropogenic input: a study case Pasărea River. *International Scientific Symposium Current Trends in Natural Science*, Book of abstract. University of Pitești: 53-54.
- POMEROY L. R. & WIEBE W. J. 2001. Temperature and substrates as interactive limiting factors for marine heterotrophic bacteria. *Aquatic Microbial Ecology*. Oregon State University Press. **23**: 187-204.
- RAJI M. I. O., IBRAHIM Y. K. E., TYTLER B. A., EHINMIDU J. O. 2015. Faecal Coliforms (FC) and Faecal Streptococci (FS) Ratio as Tool for Assessment of Water Contamination: A Case Study of River Sokoto, Northwestern Nigeria. *Handbook on the Emerging Trends in Scientific Research*. Wiley Press. London. **3**: 8-11.
- RUGINESCU R., ENACHE M., POPESCU O., GOMOIU I., COJOC ROXANA, BĂTRÎNESCU-MOTEAU C., MARIA G., DUMBRĂVICIAN M., NEAGU SIMONA. 2022. Characterization of Some Salt-Tolerant Bacterial Hydrolases with Potential Utility in Cultural Heritage Bio-Cleaning. *Microorganisms*. Cambridge University Publishing. London. **10**(3): 644.
- SELVAM S., RAVINDRAN A., VENKATRAMANAN S., SINGARAJA C. 2015. Assessment of heavy metal and bacterial pollution in coastal aquifers from SIPCOT industrial zones, Gulf of Mannar, South Coast of Tamil Nadu, India. *Applied Water Science*. Scimago Press. London. **7**(2): 897-913.
- SINTON L. W., DONNISON A. M., HASTIE C. M. 1993. Faecal streptococci as faecal pollution indicators: A review. Part II: Sanitary significance, survival, and use. *New Zealand Journal of Marine and Freshwater Research*. Taylor & Francis Publisher. London. **27**(1): 117-137.
- SINTON L. W., FINLAY R. K. HANNAH D. J. 1998. Distinguishing human from animal faecal contamination in water: A review. *New Zealand Journal of Marine and Freshwater Research*. Taylor & Francis Publisher. London. **32**(2): 323-348.
- ZARNEA G. 1994. *Tratat de microbiologie generală*. Edit. Academiei Române. București. **5**. 1078 pp.
- WANG S., GAO M., LI Z., SHE Z., WU J., ZENG D., GUO L., ZHAO Y., GAO F., WANG X. 2016. Performance evaluation, microbial enzymatic activity and microbial community of sequencing batch reactor under long-term exposure to cerium dioxide nanoparticule. *BioresourceTechnology*. Elsevier. Paris. **220**: 262-270.
- \*\*\*. APHA. 1998. Standard methods for the examination of water and wastewater. 20<sup>th</sup> ed. American Public Health Association, Washington, DC.
- \*\*\*. Council Directive 98/83/EC of 3 November 1998 on the quality of water intended for human consumption.
- \*\*\*. Standards for drinking water quality - Law 458/2002, Monitorul Oficial nr. 875 din 12 decembrie 2011.
- \*\*\*. STAS 3001-91. Apa. Analiză bacteriologică. Institutul Român de Standardizare (IRS). București: 1-32

**Podosu (Vlad) Aurelia, Neagu Simona, Lucaci Anca Ioana, Ruginescu Robert, Cojoc Roxana, Bătrînescu-Moteau Costin, Purcărea Cristina, Enache Mădălin**  
Institute of Biology Bucharest, Romanian Academy, Spl. Independenței no. 296, sect. 6, 060031, Bucharest, Romania.  
E-mails: vladaurelia24@yahoo.com; costinbatrinescu@yahoo.com; madalin.enache@ibiol.ro

Received: April 15, 2023  
Accepted: August 23, 2023